



Supplemental Materials

for

Mass Spectrometry as a Tool to Enhance "-omics" Education
Michael J. Wolyniak^{1*}, Nathan S. Reyna², Ruth Plymale², Welkin H. Pope³,
and Daniel E. Westholm⁴

¹*Department of Biology, Hampden-Sydney College, Hampden-Sydney, VA 23943*, ²*Department of Biology, Ouachita Baptist University, Arkadelphia, AR 71998*, ³*Department of Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260*, ⁴*Department of Biology, The College of St. Scholastica, Duluth, MN 55811*

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*Corresponding author. Mailing address: Department of Biology,
Hampden-Sydney College, Box 183, Hampden-Sydney, VA 23943.
Phone: 434-223-6175. E-mail: mwolyniak@hsc.edu.
Received: 27 August 2017, Accepted: 27 November 2017, Published:
16 February 2018.

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Appendix 1: Experimental protocol for mass spectrometry of *M. smegmatis* cell pellet infected by mycobacteriophage

- 1) Inoculate several cultures of *M. smegmatis* (e.g. dilute 50/100/200/400/800 μ l smeg culture into 50 ml sterile growth media-the goal is to have one culture at an OD of 0.4 the next day).
- 2) Incubate at overnight at 37°C with shaking.
- 3) Measure OD600 of cultures (set OD= to 3.5×10^7)
- 4) Take 5 ml of culture with OD=0.4 and spin down in 15 ml conical tube @ 500 \times g for 5 min.
- 5) Remove 4 ml of supernatant and resuspend pellet in remaining broth.
- 6) Infect concentrated culture with phage at an MOI=10. Mix gently.
- 7) Incubate for at 37°C for 15 min.
- 8) Bring volume of conical tube back to 5 ml with pre-warmed broth culture media.
- 9) Transfer contents to sterile 50 ml flask containing several cut pipette tips
- 10) Incubate flasks at 37°C with shaking for 3-3.5 hours.
- 11) Transfer 1.5 ml from flask to two sterile microcentrifuge tubes.
- 12) Pellet the phage-infected cells at 14,000 \times g for 5 min.
- 13) Remove supernatant. Resuspend each pellet in 750 μ l of phage buffer. Combine resuspended pellets into one tube.
- 14) Spin combined pellets at 14,000 \times g for 8 min.
- 15) Remove supernatant.
- 16) Store infected cell pellet at -80°C.

Sample Multiplicity of Infection (MOI) calculation:

The *M. smegmatis* culture OD600 spec reading gave a concentration of 2.19×10^7 cells/ml. Your phage lysate is at 6×10^9 pfu/ml. You want to infect 1 ml of bacterial cells (2.19×10^7 cells) with an MOI of 10 (10 phage particles per cell in infection).

Calculation:

$(2.19 \times 10^7 \text{ cells}) \times (10 \text{ phages/cell}) \times (1 \text{ ml} / 6 \times 10^9 \text{ pfu}) = 0.1825 \text{ ml} = 182.5 \mu\text{l}$ phage lysate for infection